Section 6 (Texas Traditional) Report Review

Attachment to letter dated OCT 10 2003

TPWD signature date on report 12/8/07

Project Title: Phylogeography of cave crickets (Ceuthophilus spp.) in Central Texas: A keystone taxon for the conservation and management of federally listed endangered cave arthropods

Final or Interim Report? Final

Grant #: E-60

Reviewer Station: Austin ESFO

Lead station was contacted and concurs with the following comments:
☐ Yes  ☐ No  ☒ Not applicable (reviewer is from lead station)

Interim Report (check one):
☐ is acceptable as is
☐ is acceptable as is, but comments below need to be addressed in the next report
☐ needs revision (see comments below)

Final Report (check one):
☒ is acceptable as is
☐ is acceptable, but needs minor revision (see comments below)
☐ needs major revision (see comments below)

Comments:
FINAL REPORT

As Required by

THE ENDANGERED SPECIES PROGRAM

TEXAS

Grant No. E - 60

Endangered and Threatened Species Conservation

Phylogeography of cave crickets (Ceuthophilus spp.) in central Texas: A keystone taxon for the conservation and management of federally endangered cave arthropods

Prepared by:

Steve Taylor

Carter Smith
Executive Director

Mike Berger
Division Director, Wildlife

5 December 2007
FINAL REPORT

STATE: Texas  GRANT NUMBER: E-60

GRANT TITLE: Phylogeography of cave crickets (Ceuthophilus spp.) in central Texas: A keystone taxon for the conservation and management of federally endangered cave arthropods

REPORTING PERIOD: 10/01/04 to 12/1/07

OBJECTIVE(S):

To examine broad spatial patterns of genetic relatedness and population structure among cave-inhabiting Ceuthophilus populations of the Edwards Plateau and Balcones Escarpment in central Texas in relation to: 1) the known distribution and biology of the species, 2) the geological context in which they occur, and 3) the genetic relatedness of other cavernicolous presently being studied by others (e.g., Hedin & Paquin), and 4) the management and regulatory issues related to terrestrial karst invertebrates in central Texas.

Summary Of Progress:

Please see Attachment A (.pdf file).

Significant Deviation:

None

Location: Several Central Texas Counties

Cost: Not available at time of this report

Prepared by: Craig Farquhar  Date: 5 December 2007

Approved by: C. Craig Farquhar  Date: 8 Dec 07
Phylogeography of cave crickets (*Ceuthophilus* spp.) in central Texas: A keystone taxon for the conservation and management of federally listed endangered cave arthropods

Steven J. Taylor¹, Jason D. Weckstein², Daniela M. Takiya³, Jean K. Krejca⁴, John D. Murdoch⁵, George Veni⁶, Kevin P. Johnson¹, and James R. Reddell⁷

¹Biodiversity & Ecological Entomology
Illinois Natural History Survey
1816 S. Oak Street
Champaign IL 61820-6953 USA

²Zoology-Bird Division
The Field Museum
1400 S. Lake Shore Drive
Chicago, IL 60605-2496 USA

³Departamento de Zoologia
Setor de Ciências Biológicas
Universidade Federal do Paraná
Caixa Postal 19020
81531-980, Curitiba, Paraná Brazil

⁴Zara Environmental, LLC
118 West Goforth Rd
Buda, TX 78610 USA

⁵Department of Genetics
Yale University School of Medicine
333 Cedar Street, P.O. Box 208005
New Haven, CT 06520-8005 USA

⁶National Cave & Karst Research Institute
1400 Commerce Dr.
Carlsbad, NM 88220-9187 USA

⁷Curator of Invertebrate Zoology
Texas Memorial Museum
The University of Texas at Austin
2400 Trinity St
Austin, TX 78705-5730 USA

Contact:
email: sjtaylor@uiuc.edu
phone: 217-649-0240

30 November 2007

prepared for:
Attn: Dr. C. Craig Farquhar
Section 6 Grant Program Coordinator, Wildlife Division
Texas Parks and Wildlife Department
4200 Smith School Road, Austin, Texas  78744 USA
Abstract

Ceuthophilus spp. (Orthoptera: Rhaphidophoridae) in two subgenera (Ceuthophilus and Geotettix) were examined phylogenetically, morphologically, and biogeographically in order to improve our understanding of these species that carry nutrients into caves and are thus closely tied to rare and endangered troglobites in central Texas. Crickets were collected from 43 caves across central Texas, and outgroup taxa were collected in west Texas, Kentucky, Missouri, New Mexico, and Mexico. We analyzed 1263 base pairs of mitochondrial DNA from the COI and ND5 genes and complemented this data with morphometric analyses of 19 morphological characters. Phylogenetic trees from the molecular analysis allowed us to select representatives from 25 clades for morphological work.

The molecular data show relatively high levels of genetic variation and phylogeographic structure. This variation is higher than that predicted by the currently accepted species level taxonomy of Ceuthophilus. These genetically divergent lineages uncovered by our DNA sequence data may be cryptic species. Members of the subgenus Ceuthophilus are known to commonly forage outside of caves leading us to predict that they might also be better at dispersing than members of the subgenus Geotettix, which are not known to leave their caves. However, contrary to our expectations, haplotype phylograms of the two subgenera indicate that the subgenus Ceuthophilus has deeper genetic structure than the subgenus Geotettix. Therefore, Ceuthophilus populations have been isolated for a longer period than Geotettix populations. Another unexpected result is that multiple haplotypes from genetically divergent groups are found in the same cave on multiple occasions, another indicator of cryptic species.

The morphological dataset included 19 discreet characters and 31 continuous characters that were used to create a morphology-based tree and plotted in a principal component analysis. The morphological and genetic trees are similar only on a rough level in that the two subgenera are generally distinct (though Geotettix is monophyletic in the genetic analysis and one morphological analysis and paraphyletic in another morphological analysis). Morphological characters are also divergent within monophyletic clades and convergent among clades, also pointing to probable undescribed species.

As Ceuthophilus spp. are a key component in central Texas karst invertebrate conservation, identifying and describing any new species is a high conservation priority. Different species may have different life history patterns, foraging behaviors, and use of caves.
Introduction

Understanding the role that cave crickets play in cave communities is important for the management and protection of endangered karst invertebrates in central Texas. Cave crickets are key species (Taylor et al. 2005, Lavoie et al. 2007) in cave communities because 1) they often occur in large numbers, and 2) they are one of the primary modes of transporting energy from the surface environment into the caves. Effective conservation and management of Texas’ federally listed endangered karst invertebrates (USFWS 1988, 1993, 2000) requires understanding the direct and indirect dependence of cave communities on the biology of Ceuthophilus cave cricket taxa. However, our knowledge of the occurrence and distribution of Texas’ cave cricket taxa remains in its infancy. Here we seek to expand our understanding of the spatial distribution of evolutionary lineages (both described and undescribed species) of central Texas Ceuthophilus species. These data can be utilized in future studies to better define the biology (food sources, annual life cycle, numbers of eggs produced, energy flow) of both the crickets and the endangered karst invertebrates.

The genus Ceuthophilus currently includes 89 valid species and geographical races in North America. The generic taxonomy and morphology of Ceuthophilus was thoroughly revised by Hubbell (1936) who studied 71 primary types and over 17,000 specimens (Strohecker 1936). He divided the genus into three subgenera: Ceuthophilus (Hemiudeopsylla) with eight species restricted to southwestern North America; Ceuthophilus (Geotetix) including 20 species mostly distributed in the Great Basin and Great Plains regions; and the nominotypical Ceuthophilus (Ceuthophilus) that includes 62 species which comprise the majority of species inhabiting forested areas of North America. Furthermore, Hubbell (1936) accommodated all species in species groups, and in turn accommodated those in series, in an attempt to express his understanding of relationships (“close alliance”) within the genus. Since Hubbell’s revision, the genus Ceuthophilus has received little attention, although various researchers recognize a number of undescribed species. For example, unpublished observations by Reddell, corroborated by Krejca and Taylor, suggested that C. (G.) cunicularis represents more than one species. Many Ceuthophilus taxa cannot be easily distinguished, and the taxonomy of the genus Ceuthophilus is in need of revision because the species descriptions of Hubbell’s (1936) monograph refer to differing sets of characters, confounding what should be fairly straightforward species identifications (Cohn 2000).

The present study is an assessment of the molecular and morphological variation of species of cavernicolous Ceuthophilus species occurring in Texas and Mexico. We examine broad spatial patterns of genetic relatedness and population structure among cave-inhabiting Ceuthophilus of the Edwards Plateau and Balcones Escarpment in central Texas in relation to the:

1) known distribution and biology of the species,
2) geological context in which they occur,
3) genetic relatedness of other cavernicoles
4) management and regulatory issues related to terrestrial karst invertebrates in central Texas.

Species definitions by Hubbell (1936) and hypotheses of relationships generated based on morphology are compared to a phylogenetic hypothesis generated based on DNA-sequence data. The primary species of interest are cave crickets typical of central Texas caves. They belong to the undescribed Ceuthophilus (C.) “species B”, Ceuthophilus (G.) cunicularis which is thought to
represent a complex of two or more taxa, not all of which have been described, and the widely spread and often abundant *Ceuthophilus (C.) secretus*.

**Cave cricket biology**

*C. secretus* (Figure 1) and other, undescribed, *C. secretus* (*C. cunicularis*) species in central Texas forage at night on the surface, returning to the cave to roost during the day. Their feces, dead bodies, and eggs (laid in the caves) constitute a significant portion of the available energy in the cave environment. Crickets which emerge from central Texas caves to forage at night are currently recognized as *C. (C.) secretus* and *C. secretus* (*C. cunicularis*) “species B” – the latter is an undescribed taxon, with unclear affinities to *C. (C.) secretus*. Recently, Taylor et al. (2003, 2004, 2005) demonstrated that *C. secretus* commonly forages more than 100 meters from cave entrances. A third species, *C. (Geotettix) cunicularis*, is also reported from many of the endangered species caves, though it rarely, if ever, leaves caves to forage.

![Figure 1. *Ceuthophilus (Ceuthophilus) secretus* from Fort Hood, Texas (Bell and Coryell counties).](image)
The life histories of endangered cave arthropods and the various (likely greater than three) species of *Ceuthophilus* are tightly interconnected. The most obvious relationship is the predator/prey relationship between the endangered *Rhadinus* beetles (Mitchell 1971) and one or more of the cricket taxa. Cave-adapted *Rhadinus* beetles (Figure 2) are known to prey upon eggs of *Ceuthophilus* species. Adult female *Ceuthophilus* spp. insert their ovipositors deeply in the substrate and deposit a single, fairly large egg. Based on more detailed studies of Rhaphidophorid crickets elsewhere (Kane and Poulson 1976, Hubbell and Norton 1978, Griffith 1990, Lavoie et al. 2007) it is likely that the crickets make a series of false oviposition holes to limit beetle depredation. One of us (JKK) has observed a female *Ceuthophilus* repeatedly inserting her ovipositor in the substrate, and studies on eastern and midwestern North American *Hadenoeus* species demonstrate an obligate relationship between the cave cricket species and their beetle predators (e.g., Poulson et al. 1995, Helf et al. 1996, Studier 1996, Lavoie et al. 2007). These beetle/cricket species pairs are unique in different geographic areas (e.g., the karst of central Kentucky versus the Appalachians). In central Texas, three species of *Rhadinus* are federally listed as endangered (*Rhadinus persephone* in Travis and Williamson counties, plus *Rhadinus exilis* and *Rhadinus infernalis* in Bexar County), and it is probable that a significant portion of their food comes in the form of cave cricket eggs.

Relationships between other federally listed endangered cave arthropods and cave crickets are less direct but clearly important. In Bell and Coryell counties (the northeastern limit of our study area), Taylor et al. (2003) documented similar spatial distributions of *Cicurina* spiders, cave-adapted springtails, and *C. (C.) secretus*. It appears that the springtails feed on the guano of roosting cave crickets (and/or the fungi and bacteria which are breaking down the cricket guano) and the predatory *Cicurina* spiders are, in turn, feeding on the springtails. Cokendolpher (personal communication 2002) successfully maintained *Cicurina* spp. in the laboratory on a diet of cave springtails, corroborating portions of the above relationships. In addition, one of us (JRR) observed both pseudoscorpions (*Tartarocreagris* spp.) and troglobitic *Cicurina* spp. spiders feeding on *Ceuthophilus* spp. nymphs.

The relationship between other endangered cave arthropods (e.g., *Batrisodes* spp.) and the cave crickets in central Texas is less clear, but cave crickets, and their feces and eggs, provide one of the most important sources of energy driving the cave ecosystem (excluding a few caves dominated by colonial bats or others which regularly flood) (Taylor et al. 2003, 2004, 2007). Poulson (1992) reviewed in detail the better-studied terrestrial cave communities of Mammoth Cave, Kentucky, and
discussed demonstrable links between the rhaphidophorid *Hadenoecus subterraneus* and various cave-adapted invertebrates.

*Ceuthophilus* biology and distribution in Bell and Coryell counties has been shown to differ between the more vagile *C. (C.) secretus* and the seemingly cave-limited crickets presently referred to as *C. (G.) cunicularis* (Taylor et al. 2003). *Ceuthophilus (C.) secretus* leave these caves at night to forage during the warmer months, sometimes traveling more than 100 m from a cave entrance, and occasionally moving between caves (Taylor et al. 2004). In another similarly mobile cave cricket species, *Ceuthophilus (Ceuthophilus) gracilipes*, Cockley et al. (1977) found relatively low levels of genetic variation within and between populations. At the other extreme, *C. (G.) cunicularis* was never observed leaving caves during studies by Taylor et al. (2003, 2005, 2007) and was only once been observed exiting a cave during extensive monitoring at Camp Bullis in Bexar County (Veni and Associates 2006). Within caves, *C. (C.) secretus* are typically found on bedrock ceilings whereas *C. (G.) cunicularis* are typically found on the walls and floors (Taylor et al. 2003). Finally, carbon and nitrogen stable isotope data (Taylor et al. 2004, 2007) suggest that *C. (C.) secretus* and *C. (G.) cunicularis* forage on different foods and may even occupy differing trophic levels. Collectively, these differences in ecology and mobility may be reflected in levels of population genetic structure, as has been found in other cavernicolous arthropods (Caccone 1985, Caccone and Sbordoni 1987).

**Endangered Species Management Implications**

The management of the federally listed endangered terrestrial cave invertebrates requires an understanding of the phylogenetic relationships and population genetic structure of the cave crickets upon which they depend. Different cricket species, whether named or merely identified as genetically unique entities, may well have differing life history strategies that have implications for the federally listed endangered terrestrial troglobites in central Texas. The foraging range of these crickets is a crucial piece of information for effective management of cave and karst ecosystems. For example, land managers may wish to control access to, or control red imported fire ant (RIFA, *Solenopsis invicta*) populations in all or part of the foraging range of the crickets to help maintain natural surface and cave communities. The obvious differences in microhabitat utilization by *C. (C.) secretus* and *C. (G.) cunicularis* documented in Bell and Coryell counties (Taylor et al. 2003) may not be applicable to populations in Travis/Williamson or Bexar counties if, as it is strongly suspected, the *C. (G.) cunicularis* in Bell and Coryell counties is actually an undescribed species. Making management recommendations with regard to cave crickets within the range of the endangered species caves depend on knowing what cricket taxa are present, because the biology of unstudied taxa may differ from related taxa in different areas. Also, patterns of genetic variation in cave crickets may be useful for identifying areas of unique biological diversity.

**Materials and Methods**

We collected mitochondrial DNA (mtDNA) sequence data to compare patterns of genetic structure between the three cricket species across their geographic ranges. Mitochondrial sequences have been useful for comparing levels of population genetic structure in other cavernicoles (Caccone and Sbordoni 2001). In total we sequenced mtDNA from 309 individual cave crickets.
Sampling locations and collection methods

The geographic area encompassed by this study focus primarily on counties within the Balcones Escarpment and Edwards Plateau (Figure 3). The federally listed endangered cave invertebrates are confined to the Balcones Escarpment within Bexar, Travis, and Williamson counties, with a few localities in eastern Burnet County. Cave crickets in the subgenera *Geotettix* and *Ceuthophilus* are codistributed across central Texas. We sampled multiple individuals from each subgenus at each cave where possible, with *Ceuthophilus* specimens from 43 caves distributed across 20 Texas counties and two caves in Mexico. We also obtained specimens from caves in Kentucky (*Hadenoecus subterraneus*), Missouri (*Ceuthophilus [Ceuthophilus] gracilipes, Ceuthophilus [Ceuthophilus] williamsoni*), and New Mexico (*Ceuthophilus [Ceuthophilus] longipes, Ceuthophilus [Geotettix] carlsbadensis, and Ceuthophilus [Ceuthophilus] conicaudus*) to serve as outgroup taxa (Figure 3), for a total of 50 sampling sites (Table 1). These sample sites were selected to maximize our understanding of potential barriers to gene flow. We suspect that major geological discontinuities (Figure 4), rivers (Figure 5) or ecoregional differences (Figure 6) could serve as barriers to gene flow.

Figure 3. Distribution of all sites from which Rhaphidophoridae (Orthoptera) were sampled for this study, including outgroup taxa.
Table 1. Names and county-level locations for sites sampled for this study.

<table>
<thead>
<tr>
<th>Country</th>
<th>State</th>
<th>County</th>
<th>Caves</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mexico</td>
<td>Coahuila</td>
<td></td>
<td>Cueva de La Azufrosa, Cueva de Casa Blanca</td>
</tr>
<tr>
<td>USA</td>
<td>Kentucky</td>
<td>Barren</td>
<td>Dogwood Cave</td>
</tr>
<tr>
<td></td>
<td>Missouri</td>
<td>Pulaski</td>
<td>Breeden Cave, Davis Cave #3</td>
</tr>
<tr>
<td>New Mexico</td>
<td>Eddy</td>
<td></td>
<td>Carlsbad Cavern (&amp; outside of cave)</td>
</tr>
<tr>
<td>Texas</td>
<td>Bandera</td>
<td></td>
<td>near Haby Salamander Cave</td>
</tr>
<tr>
<td></td>
<td>Bexar</td>
<td></td>
<td>MARS Shaft, Poor Boy Baculum Cave, Robber</td>
</tr>
<tr>
<td></td>
<td>Brewster</td>
<td></td>
<td>400 Foot Cave</td>
</tr>
<tr>
<td></td>
<td>Comal</td>
<td></td>
<td>Camp Bullis Bat Cave, Camp Bullis Cave No. 1, Preserve Cave, Temple of Doom</td>
</tr>
<tr>
<td></td>
<td>Coryell</td>
<td></td>
<td>Mixmaster Cave, Rocket River Cave</td>
</tr>
<tr>
<td></td>
<td>Edwards</td>
<td></td>
<td>Deep Cave, Devils Sinkhole, Punkin Cave, Writing on the Rocks Cave</td>
</tr>
<tr>
<td></td>
<td>Hays</td>
<td></td>
<td>Ezell's Cave</td>
</tr>
<tr>
<td></td>
<td>Kendall</td>
<td></td>
<td>Dead Man's Cave</td>
</tr>
<tr>
<td></td>
<td>Kerr</td>
<td></td>
<td>Schroeder Bat Cave</td>
</tr>
<tr>
<td></td>
<td>Kinney</td>
<td></td>
<td>Kickapoo Cavern</td>
</tr>
<tr>
<td></td>
<td>Mason</td>
<td></td>
<td>Behren's Grotto, Porcupine Pit, Swift Cave</td>
</tr>
<tr>
<td></td>
<td>Medina</td>
<td></td>
<td>Ground Hog Cave</td>
</tr>
<tr>
<td></td>
<td>Pecos</td>
<td></td>
<td>Amazing Maze Cave</td>
</tr>
<tr>
<td></td>
<td>Real</td>
<td></td>
<td>All the Wonders and Joys Cave, Little Dry Frio</td>
</tr>
<tr>
<td></td>
<td>San Saba</td>
<td></td>
<td>(a cave without any name), Cicurina Cave, Lemons Ranch Cave, Puberty Pit, Rattlesnake Drop, Turtle Shell Cave</td>
</tr>
<tr>
<td></td>
<td>Sutton</td>
<td></td>
<td>Caverns of Sonora, IH-10 Cave,</td>
</tr>
<tr>
<td></td>
<td>Travis</td>
<td></td>
<td>Lamm Cave, Lost Oasis Cave, Testudo Tube</td>
</tr>
<tr>
<td></td>
<td>Uvalde</td>
<td></td>
<td>Finley Bat Cave</td>
</tr>
<tr>
<td></td>
<td>Val Verde</td>
<td></td>
<td>Big Tree Cave, (cave near Del Rio), Fern Cave</td>
</tr>
<tr>
<td></td>
<td>Williamson</td>
<td></td>
<td>Temples of Thor</td>
</tr>
</tbody>
</table>
Figure 4. Distribution of sites in Texas, New Mexico, and Mexico from which *Ceuthophilus* spp. (Orthoptera: Rhaphidophoridae) were obtained for this study, overlaid on the distribution of karst (not including Mexico). Karst map modified after Veni and Duchene (2001).
Figure 5. Distribution of sites in Texas, New Mexico, and Mexico from which *Ceuthophilus* spp. (Orthoptera: Rhaphidophoridae) were obtained for this study, overlaid on the major streams and rivers of Texas.
Figure 6. Distribution of study sites in Texas, New Mexico, and Mexico from which *Ceuthophilus* spp. (Orthoptera: Rhaphidophoridae) were obtained for this study, overlaid on US EPA Level III and IV Ecoregions of Texas (Griffith et al. 2004). Level III Ecoregions indicated by colors and bold black text, with selected Level IV regions outlined in white, with white text. Chihuahuan Deserts: a. Chihuahuan Basins and Playas, b. Chihuahuan Montane Woodlands; Southern Texas Plains: c. Semiarid Edwards Bajada; Edwards Plateau: d. Semiarid Edwards Plateau, e. Edwards Plateau Woodland, f. Llano Uplift, g. Balcones Canyonlands; Cross Timbers: h. Limestone Cut Plain.
especially in the largely cave-limited *C. (G.) cunicularis*. Because *C. (C.) secretus* is able to travel over land for significant distances (more than 100 meters over short periods of time [Taylor et al. 2003, 2004, 2005]) and can move between karst features (Taylor et al. 2004), higher levels of gene flow may be evident in this species.

**Collecting methods used and tissue sampling**

Specimens were captured by hand and placed in 95% ethanol to preserve DNA. An effort was made to collect several adult individuals of both sexes and both subgenera at each site. In the lab, a single leg was removed from each specimen for use in DNA extraction. Selected specimens were used for morphological analysis in this study, and these, along with the remaining portion of each specimen will be deposited in the insect collection of the Illinois Natural History Survey as vouchers available for future analyses. Unlike the federally list endangered karst invertebrates, the cave crickets usually occur in sufficiently large numbers (sometimes thousands of individuals in a single cave) that collecting 3-5 individuals from a single cave on a one-time basis will have an inconsequential effect on the population size.

**Sample sizes and localities to be samples**

We collected 1263 base pairs (bp) of mitochondrial DNA (mtDNA) sequence data (COI and ND5 regions) from 122 individual cave crickets in the subgenus *Geotettix*, including *C. (G.) cunicularis*, *Ceuthophilus (Geotettix) polingi*, and 179 individual cave crickets in the subgenus *Ceuthophilus* including *C. (C.) secretus*, *C. (C.) conicaudus*, and the undescribed taxon *C. (C.) “species B.”* We also collected mtDNA sequence data from the following outgroup taxa to root phylogenetic trees in some analyses: *H. subterranean* (*n* = 1), *C. (C.) gracilipes* (*n* = 1), *C. (C.) longipes* (*n* = 2), *C. (G.) carlsbadensis* (*n* = 3), and *C. (C.) williamsoni* (*n* = 1).

**Extraction, PCR, and DNA sequencing**

We extracted whole genomic DNA from a single leg of each specimen using a Dneasy extraction kit following the kit protocol for animal tissues (Qiagen, Valencia, California). For each specimen we sequenced 1263 bp of mitochondrial DNA (mtDNA) including 850 bp of Cytochrome Oxidase I (COI) and 413 bp of nicotinamide adenine dinucleotide dehydrogenase subunit 5 (ND5). We amplified an 850 bp fragment of COI using the primers C1-J-1718 (Simon et al. 1994) and H7005 (Hafner et al. 1994) and sequenced this fragment using the C1-7-1718, H7005, and two of three internal primers designed specifically for this study: CeuthCOI (5’-GATCCTGCTGGAGGAGATCC-3’), and either CeuthCOIH (5’-GAATTGGATCTCTCCACCACGAGG-3’) or CeuthCOIHcunn (5’-GAATTGGATCTCTCCACGAGGAGG-3’). We amplified and sequenced a 413 bp fragment of ND5 using the primers F7081 and R7495 (Yoshizawa 2004). For COI PCR amplifications we used the following thermal cycling profile: 94°C for 2 min, 35 cycles of 94°C for 30s, 46°C for 30s, 72°C for 1 min, and 72°C for 7 min. Whereas for ND5 PCR amplifications we used the following thermal cycling profile: 94°C for 2 min, 35 cycles of 94°C for 30s, 42°C for 30s, 65°C for 30s, and 65°C for 7 min. We verified all PCR products on a 1% agarose gel and purified these PCR products using a QIAquick PCR Purification kit (Qiagen, Valencia, California).

Cycle sequencing reactions were performed at the University of Illinois DNA sequencing facility using an ABI Big Dye kit (Applied Biosystems, Foster City, California), the above-listed
primers, and approximately 75 ng of purified PCR product. We ran purified sequencing reaction products on an ABI 3730 capillary electrophoresis system (Applied Biosystems) and used Sequencher (ver. 4.5, GeneCodes Co., Ann Arbor, Michigan) to reconcile double-stranded sequences and to align sequences for analysis.

Phylogenetic and population genetic analyses

We used MacClade (version 4.05; Maddison and Maddison 2002) to collapse the entire dataset down to unique sequences (haplotypes) for phylogenetic analysis. We reconstructed separate phylogenetic trees for the subgenera *Geotettix* and *Ceuthophillus* using Neighbor-Joining analysis as implemented in the computer programs PAUP* (ver. 4.0beta10; Swofford 2001) and used the outgroups listed above to root the topologies. We also calculated uncorrected pairwise sequence divergence (p-distance) using PAUP* (Swofford 2001) between taxa in these two subgeneric clades to compare the relative levels of divergence within each of the subgenera.

A reduced taxon dataset was used to perform a Bayesian phylogenetic analysis as implemented in the computer program MrBayes (ver. 3.1.1; Ronquist and Huelsenbeck 2003). For the Bayesian analysis we analyzed the COI and ND5 mtDNA sequences for forty-seven terminal taxa using a mixed-model approach by partitioning each gene by codon position (total of 6 partitions). The analysis was conducted with two independent runs of 5,000,000 generations each with 4 mcmc chains. A majority-rule consensus with average branch lengths and posterior probabilities was constructed based on 10,854 distinct trees after a 1,000,000 generation burn-in (trees sampled prior to the analysis reaching stationarity) was discarded (Leaché and Reeder 2002).

To assess whether there was an influence of geography on the phylogenetic results, the resulting Bayesian tree for the *Ceuthophilus* (*Geotettix*) subgenus was divided into clades containing one to four of the previously defined genetic lineages. This tree was then plotted onto a map using geographical coordinates of the specimens (terminal taxa) with the module Cartographer of Mesquite (Maddison & Maddison 2006a, 2006b). We also compared this tree topology to the morphological data and used it to identify matching pairs of males and females for morphological analysis.

Taxon sampling for morphological study

For purposes of reference throughout this report, we named monophyletic lineages, or clades, of crickets using letters that were arbitrarily assigned based on the molecular phylogeny. These letter designations do not necessarily imply species breaks, nor do they relate to terminology used by other authors and researchers for undescribed species (e.g., *Ceuthophilus* “species B”). The *Ceuthophilus* included in this study are divided into two main groups, corresponding to their respective subgenera: (1) monophyletic lineages A-J, *C. (C.) gracilipes*, *C. (C.) conicaudus*, and *C. (C.) secretus* (belonging to nominative subgenus *Ceuthophilus*); and (2) monophyletic lineages K-T, *C. (G.) carlsbadensis*, *C. (G.) cunicularis*, and *C. (G.) polingi* (all belonging to subgenus *Geotettix*). Morphological characters were studied from adult male and female specimens from each lineage (see specimen list in Table 1). The morphology of lineage I was not studied because only immature specimens were available, and lineage Q was further divided into Q and Q2. Voucher numbers and geographical origins are listed in Table 2.
Table 2. Voucher numbers and Texas county of origin (except for Mexico [MX] state, New Mexico [NM] and Missouri [MO] counties) of adult *Ceuthophilus* specimens for each molecular lineage which were studied for the morphological analysis.

<table>
<thead>
<tr>
<th>Subgenus</th>
<th>Molecular Lineage (Clade)</th>
<th>Male</th>
<th>Female</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>C. (Ceuthophilus)</em> spp.</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>A</td>
<td>-</td>
<td>448: Sutton</td>
<td></td>
</tr>
<tr>
<td>B</td>
<td>-</td>
<td>310: Travis</td>
<td></td>
</tr>
<tr>
<td>C</td>
<td>083: Comal</td>
<td>140: Brewster</td>
<td></td>
</tr>
<tr>
<td>D</td>
<td>444: Medina</td>
<td>443: Medina</td>
<td></td>
</tr>
<tr>
<td>E</td>
<td>459: Val Verde</td>
<td>460: Val Verde</td>
<td></td>
</tr>
<tr>
<td>F</td>
<td>-</td>
<td>318: Williamson</td>
<td></td>
</tr>
<tr>
<td>G</td>
<td>-</td>
<td>215: Bexar</td>
<td></td>
</tr>
<tr>
<td>H</td>
<td>-</td>
<td>452: Uvalde</td>
<td></td>
</tr>
<tr>
<td>J</td>
<td>-</td>
<td>139: Brewster</td>
<td></td>
</tr>
<tr>
<td>C. (C.) <em>conicaudus</em></td>
<td>258: Eddy (NM)</td>
<td>259: Eddy (NM)</td>
<td></td>
</tr>
<tr>
<td>C. (C.) <em>gracilipes</em></td>
<td>009: Pulaski (MO)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C. (C.) <em>longipes</em></td>
<td>-</td>
<td>253: Eddy</td>
<td></td>
</tr>
<tr>
<td>C. (C.) <em>secretus</em></td>
<td>229: Travis</td>
<td>228: Travis</td>
<td></td>
</tr>
<tr>
<td><em>C. (Geotettix)</em> spp.</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>K</td>
<td>038: Edwards</td>
<td>043: Edwards</td>
<td></td>
</tr>
<tr>
<td>L</td>
<td>078: Val Verde</td>
<td>336: Edwards</td>
<td></td>
</tr>
<tr>
<td>L</td>
<td>079: Val Verde</td>
<td></td>
<td></td>
</tr>
<tr>
<td>M</td>
<td>240: Bexar</td>
<td>219: Bexar</td>
<td></td>
</tr>
<tr>
<td>N</td>
<td>323: Williamson</td>
<td>324: Williamson</td>
<td></td>
</tr>
<tr>
<td>O</td>
<td>463: Val Verde</td>
<td>461: Val Verde</td>
<td></td>
</tr>
<tr>
<td>P</td>
<td>-</td>
<td>425: Bexar</td>
<td></td>
</tr>
<tr>
<td>Q</td>
<td>446: Medina</td>
<td>116: Kerr</td>
<td>114: Kerr</td>
</tr>
<tr>
<td>Q2</td>
<td>138: Brewster</td>
<td>143: Brewster</td>
<td></td>
</tr>
<tr>
<td>R</td>
<td>454: Uvalde</td>
<td></td>
<td></td>
</tr>
<tr>
<td>S</td>
<td>366: Coahuila (MX)</td>
<td>385: Coahuila (MX)</td>
<td></td>
</tr>
<tr>
<td>T</td>
<td>257: Eddy (NM)</td>
<td>256: Eddy (NM)</td>
<td></td>
</tr>
<tr>
<td>C. (G.) <em>carlsbadensis</em></td>
<td>220: Bexar</td>
<td>241: Bexar</td>
<td></td>
</tr>
<tr>
<td>C. (G.) <em>cunicularis</em> (Q)</td>
<td>148: Mason</td>
<td>153: Mason</td>
<td></td>
</tr>
<tr>
<td>C. (G.) <em>polingi</em> (R)</td>
<td>290: Coahuila (MX)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Morphological discrete character sampling

Our morphological data matrix for 25 terminal taxa, including eight male terminalia characters and 11 female external characters (Table 3, Appendix 1). These discrete characters consisted of those related to structural shape and serially homologous structures, such as spurs and denticulations. We plotted histograms of the variation observed (bin size = 10 when range was <50 and 15 when range >50) in those characters involving counts (characters 15-19), and states were delimited according to consistent breaks in the continuity of the count classes. Some character states are illustrated (Figure 7) and character and state descriptions are listed in Appendix 1. Analysis of the count data was not weighed according to the relative magnitude of the states. For *C. (C.) longipes* and seven molecular lineages, no males were available and those related characters were scored as missing data (“?”) in the matrix.

Table 3. Data matrix of discrete morphological characters of Texas *Ceuthophilus*. List of character and states are presented in Appendix 1 and illustrations of these characters in Figure 4.

<table>
<thead>
<tr>
<th>Clade</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
<th>10</th>
<th>11</th>
<th>12</th>
<th>13</th>
<th>14</th>
<th>15</th>
<th>16</th>
<th>17</th>
<th>18</th>
<th>19</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>B</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>C</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>-</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>2</td>
</tr>
<tr>
<td>D</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>-</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>E</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>-</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>3</td>
<td>4</td>
</tr>
<tr>
<td>F</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
</tr>
<tr>
<td>C. (C.) secretus</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>-</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>3</td>
<td>4</td>
</tr>
<tr>
<td>G</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>H</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>C. (C.) conicaudus</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>-</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>2</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>J</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>4</td>
</tr>
<tr>
<td>C. (G.) carlsbadensis</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>2</td>
<td>0</td>
<td>-</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td>5</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>K</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>1</td>
</tr>
<tr>
<td>L</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>3</td>
<td>1</td>
</tr>
<tr>
<td>M</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>N</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>2</td>
<td>2</td>
<td>1</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>O</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>4</td>
<td>3</td>
<td>4</td>
<td>4</td>
<td>1</td>
</tr>
<tr>
<td>P</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>C. (G.) cunicularis (Q)</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>Q2</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>2</td>
<td>2</td>
<td>3</td>
<td>4</td>
<td>1</td>
</tr>
<tr>
<td>C. (G.) polingi (R)</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>-</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td></td>
<td></td>
</tr>
<tr>
<td>S</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>T</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>3</td>
<td>2</td>
<td>2</td>
<td>3</td>
<td>0</td>
</tr>
<tr>
<td>C. (C.) longipes</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>3</td>
</tr>
<tr>
<td>C. (C.) gracilipes</td>
<td>1</td>
<td>1</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>-</td>
<td>0</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>0</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td>?</td>
<td></td>
</tr>
</tbody>
</table>
Figure 7. Morphological structures used for defining discrete characters for the phylogenetic analysis of *Ceuthophilus* species. A-F: male terminalia; A: terminal abdominal segments, lateral view, tergites labeled VIII-X; B: subgenital plates (SP), ventral view; C: epiproct (EP) and paraprocts (PP), caudal view; D: pseudosternite dorsum (PSD) with thickened dorsal rim of arch (DRA), dorsocaudal view; E: pseudosternite dorsum with pair of parallel crests (PDC); F: internal wall or arch (IWA) with pair of thickened folds, caudal view; G-J: female appendages; G: profemur with anteroventral row of setae (PAV), anterior view; H: metafemur with dorsal denticulations (DD), genicular lobe denticle (GLD), and ventral carina denticulations (VLD), anterior view; I: metatarsomere I (MT1) without plantar setae, lateral view; J: metatarsomere I (MT1) with row of plantar setae (PS), lateral view.
Morphological continuous character sampling

Continuous data for female structural characters were measured based on photographs taken of the specimens. Thirty-one measurements of the external morphology were taken for each specimen. Most of these measurements were previously used by Hubbell (1936) in species descriptions, including six diagnostic ones, which he called “primary dimensions.” Some of these measurements were posteriorly discarded because they appeared correlated ($R^2 > 0.90$) with another (e.g. length of PD vs. AD calcar, length of protarsomere I vs. metatarsomere I, length of metafemur vs. metatibia, interocular distance vs. infraocular distance, etc.). The 17 remaining measurements were used in a principal components analysis (PCA) run in Systat 10.0 (SPSS, Inc.). Representative measurements (Figure 8) measurements were reduced to two factors, which explained >85% of the total variation. Three separate analyses were run, one including all the lineages studied (Table 2), and two others including only representatives of each of the two main clades. PCA scores for each individual cricket were plotted in 2-dimensional graphs to allow a graphical visualization of the lineages studied based on their measurements.

Finally, an average linkage cluster was produced using pairwise Euclidean distances between the component scores of each taxon. This cluster analysis was compared with the molecular phylogenetic hypotheses.

Phylogenetic analysis of morphological data

The independent measurements were divided by the pronotum length of each specimen, to correct for variations in body size – the pronotal length is considered a more accurate measure of body size than overall length (Hubbell 1936), resulting 16 ratios (Table 4, Appendix 2). These ratios were then used to reconstruct a phylogenetic hypothesis in Tree analysis using New Technology (TNT, Goloboff 1999), which allows the combined parsimony analysis of continuous and discrete characters. Exact tree searches were conducted using implicit enumeration and the most parsimonious trees condensed into a strict consensus tree. Parsimony bootstrap values were also calculated.

Figure 8 (on following page). Morphological measurements taken from *Ceuthophilus* species. A: head and thorax, lateral view; PL: pronotum length; DPL: distal segment of maxillary palpus; B: face, frontal view; EA: eye area; IOL: interocular distance; C: face, lateral view; FD: fastigium distance to mesal margin of antennal fossa; D: profemur, anterior view; PFL: profemur length; E: profemur distal end, anterior view; PAV: distalmost setae of the anteroventral row length; F: mesofemur, dorsal view; GSL: posterior genicular spur length; G: metafemur, anterior view; MFW: femur largest width; MFL: femur length; H: tibial apex and metatarsus, anterior view; SDL: tibial subdistal spur length; ADC: tibial anteodorsal calcar length; MTL: tarsomere I length; I: metatarsus apex, anterior view; TCL: tarsal claw length; J: cercus, lateral view; CL: cercus length; K: ovipositor, lateral view; VVL: ventral valve length; VVA: ventral valve armed.
Table 4. Data matrix of continuous characters of female *Ceuthophilus*. Corresponding number for each ratio is listed in Appendix 2 and illustrations of measurements are presented in Figure 5.

<table>
<thead>
<tr>
<th>Taxon/Ratio</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
<th>10</th>
<th>11</th>
<th>12</th>
<th>13</th>
<th>14</th>
<th>15</th>
<th>16</th>
</tr>
</thead>
<tbody>
<tr>
<td>A</td>
<td>0.18</td>
<td>0.45</td>
<td>0.02</td>
<td>0.65</td>
<td>1.56</td>
<td>0.21</td>
<td>0.19</td>
<td>2.99</td>
<td>0.87</td>
<td>0.24</td>
<td>0.52</td>
<td>0.64</td>
<td>0.17</td>
<td>1.27</td>
<td>2.25</td>
<td>0.27</td>
</tr>
<tr>
<td>B</td>
<td>0.14</td>
<td>0.34</td>
<td>0.04</td>
<td>0.69</td>
<td>1.77</td>
<td>0.20</td>
<td>0.17</td>
<td>3.27</td>
<td>0.76</td>
<td>0.28</td>
<td>0.49</td>
<td>0.71</td>
<td>0.16</td>
<td>1.13</td>
<td>2.19</td>
<td>0.31</td>
</tr>
<tr>
<td>C</td>
<td>0.15</td>
<td>0.37</td>
<td>0.03</td>
<td>0.69</td>
<td>1.67</td>
<td>0.18</td>
<td>0.15</td>
<td>3.00</td>
<td>0.72</td>
<td>0.22</td>
<td>0.56</td>
<td>0.70</td>
<td>0.17</td>
<td>1.17</td>
<td>2.50</td>
<td>0.40</td>
</tr>
<tr>
<td>D</td>
<td>0.20</td>
<td>0.32</td>
<td>0.03</td>
<td>0.67</td>
<td>1.63</td>
<td>0.18</td>
<td>0.19</td>
<td>3.19</td>
<td>0.77</td>
<td>0.21</td>
<td>0.52</td>
<td>0.67</td>
<td>0.14</td>
<td>1.23</td>
<td>2.19</td>
<td>0.40</td>
</tr>
<tr>
<td>E</td>
<td>0.18</td>
<td>0.37</td>
<td>0.02</td>
<td>0.62</td>
<td>1.62</td>
<td>0.18</td>
<td>0.19</td>
<td>3.16</td>
<td>0.74</td>
<td>0.23</td>
<td>0.55</td>
<td>0.63</td>
<td>0.11</td>
<td>1.20</td>
<td>2.28</td>
<td>0.42</td>
</tr>
<tr>
<td>F</td>
<td>0.17</td>
<td>0.35</td>
<td>0.03</td>
<td>0.64</td>
<td>1.83</td>
<td>0.18</td>
<td>0.16</td>
<td>3.27</td>
<td>0.73</td>
<td>0.25</td>
<td>0.60</td>
<td>0.74</td>
<td>0.16</td>
<td>1.27</td>
<td>2.48</td>
<td>0.25</td>
</tr>
<tr>
<td>C. (C.) secretus</td>
<td>0.17</td>
<td>0.33</td>
<td>0.05</td>
<td>0.62</td>
<td>1.56</td>
<td>0.22</td>
<td>0.20</td>
<td>3.24</td>
<td>0.84</td>
<td>0.24</td>
<td>0.64</td>
<td>0.66</td>
<td>0.12</td>
<td>1.09</td>
<td>2.15</td>
<td>0.34</td>
</tr>
<tr>
<td>G</td>
<td>0.18</td>
<td>0.35</td>
<td>0.04</td>
<td>0.56</td>
<td>1.67</td>
<td>0.16</td>
<td>0.20</td>
<td>3.48</td>
<td>0.84</td>
<td>0.23</td>
<td>0.57</td>
<td>0.76</td>
<td>0.20</td>
<td>1.23</td>
<td>2.12</td>
<td>0.32</td>
</tr>
<tr>
<td>H</td>
<td>0.14</td>
<td>0.40</td>
<td>0.02</td>
<td>0.78</td>
<td>1.90</td>
<td>0.21</td>
<td>0.22</td>
<td>3.39</td>
<td>0.77</td>
<td>0.24</td>
<td>0.59</td>
<td>0.85</td>
<td>0.17</td>
<td>1.44</td>
<td>2.27</td>
<td>0.37</td>
</tr>
<tr>
<td>C. (C.) conicaudus</td>
<td>0.14</td>
<td>0.39</td>
<td>0.02</td>
<td>0.63</td>
<td>1.44</td>
<td>0.23</td>
<td>0.17</td>
<td>3.10</td>
<td>0.94</td>
<td>0.21</td>
<td>0.50</td>
<td>0.60</td>
<td>0.16</td>
<td>1.24</td>
<td>1.85</td>
<td>0.34</td>
</tr>
<tr>
<td>J</td>
<td>0.14</td>
<td>0.34</td>
<td>0.05</td>
<td>0.44</td>
<td>1.80</td>
<td>0.10</td>
<td>0.12</td>
<td>3.02</td>
<td>0.61</td>
<td>0.14</td>
<td>0.27</td>
<td>0.64</td>
<td>0.17</td>
<td>0.86</td>
<td>3.04</td>
<td>0.46</td>
</tr>
<tr>
<td>C. (G) carlsbadensis</td>
<td>0.10</td>
<td>0.36</td>
<td>0.03</td>
<td>0.50</td>
<td>1.44</td>
<td>0.09</td>
<td>0.09</td>
<td>2.61</td>
<td>0.58</td>
<td>0.20</td>
<td>0.31</td>
<td>0.52</td>
<td>0.19</td>
<td>1.03</td>
<td>1.39</td>
<td>0.37</td>
</tr>
<tr>
<td>K</td>
<td>0.09</td>
<td>0.33</td>
<td>0.02</td>
<td>0.55</td>
<td>1.42</td>
<td>0.12</td>
<td>0.11</td>
<td>2.35</td>
<td>0.63</td>
<td>0.23</td>
<td>0.35</td>
<td>0.50</td>
<td>0.16</td>
<td>1.07</td>
<td>1.24</td>
<td>0.26</td>
</tr>
<tr>
<td>L</td>
<td>0.08</td>
<td>0.34</td>
<td>0.01</td>
<td>0.50</td>
<td>1.32</td>
<td>0.10</td>
<td>0.12</td>
<td>2.40</td>
<td>0.59</td>
<td>0.20</td>
<td>0.31</td>
<td>0.53</td>
<td>0.20</td>
<td>0.98</td>
<td>1.22</td>
<td>0.31</td>
</tr>
<tr>
<td>M</td>
<td>0.07</td>
<td>0.34</td>
<td>0.02</td>
<td>0.60</td>
<td>1.34</td>
<td>0.12</td>
<td>0.10</td>
<td>2.22</td>
<td>0.61</td>
<td>0.19</td>
<td>0.33</td>
<td>0.54</td>
<td>0.17</td>
<td>1.18</td>
<td>1.26</td>
<td>0.32</td>
</tr>
<tr>
<td>N</td>
<td>0.06</td>
<td>0.34</td>
<td>0.04</td>
<td>0.48</td>
<td>1.32</td>
<td>0.12</td>
<td>0.12</td>
<td>2.17</td>
<td>0.59</td>
<td>0.17</td>
<td>0.36</td>
<td>0.51</td>
<td>0.18</td>
<td>1.02</td>
<td>1.14</td>
<td>0.26</td>
</tr>
<tr>
<td>O</td>
<td>0.09</td>
<td>0.36</td>
<td>0.03</td>
<td>0.53</td>
<td>1.60</td>
<td>0.09</td>
<td>0.14</td>
<td>2.63</td>
<td>0.60</td>
<td>0.26</td>
<td>0.38</td>
<td>0.55</td>
<td>0.21</td>
<td>1.19</td>
<td>1.11</td>
<td>0.29</td>
</tr>
<tr>
<td>P</td>
<td>0.08</td>
<td>0.19</td>
<td>0.02</td>
<td>0.57</td>
<td>1.41</td>
<td>0.14</td>
<td>0.09</td>
<td>2.49</td>
<td>0.66</td>
<td>0.17</td>
<td>0.28</td>
<td>0.55</td>
<td>0.18</td>
<td>1.09</td>
<td>1.28</td>
<td>0.33</td>
</tr>
<tr>
<td>C. (G) cunicularis</td>
<td>0.07</td>
<td>0.35</td>
<td>0.02</td>
<td>0.53</td>
<td>1.33</td>
<td>0.12</td>
<td>0.12</td>
<td>2.22</td>
<td>0.64</td>
<td>0.14</td>
<td>0.33</td>
<td>0.53</td>
<td>0.16</td>
<td>0.98</td>
<td>1.32</td>
<td>0.34</td>
</tr>
<tr>
<td>Q2</td>
<td>0.08</td>
<td>0.34</td>
<td>0.02</td>
<td>0.52</td>
<td>1.47</td>
<td>0.16</td>
<td>0.14</td>
<td>2.34</td>
<td>0.62</td>
<td>0.28</td>
<td>0.40</td>
<td>0.50</td>
<td>0.17</td>
<td>1.20</td>
<td>1.13</td>
<td>0.25</td>
</tr>
<tr>
<td>C. (G) polingi</td>
<td>0.08</td>
<td>0.29</td>
<td>0.02</td>
<td>0.48</td>
<td>1.27</td>
<td>0.14</td>
<td>0.12</td>
<td>2.30</td>
<td>0.67</td>
<td>0.21</td>
<td>0.36</td>
<td>0.51</td>
<td>0.16</td>
<td>0.97</td>
<td>1.18</td>
<td>0.34</td>
</tr>
<tr>
<td>S</td>
<td>0.11</td>
<td>0.54</td>
<td>0.05</td>
<td>0.50</td>
<td>2.40</td>
<td>0.19</td>
<td>0.17</td>
<td>3.95</td>
<td>0.95</td>
<td>0.28</td>
<td>0.48</td>
<td>0.89</td>
<td>0.25</td>
<td>1.50</td>
<td>1.96</td>
<td>0.57</td>
</tr>
<tr>
<td>T</td>
<td>0.07</td>
<td>0.38</td>
<td>0.03</td>
<td>0.58</td>
<td>1.55</td>
<td>0.14</td>
<td>0.09</td>
<td>2.59</td>
<td>0.63</td>
<td>0.22</td>
<td>0.32</td>
<td>0.49</td>
<td>0.21</td>
<td>1.23</td>
<td>1.16</td>
<td>0.28</td>
</tr>
<tr>
<td>C. (C) longipes</td>
<td>0.13</td>
<td>0.41</td>
<td>0.02</td>
<td>0.72</td>
<td>2.09</td>
<td>-</td>
<td>0.14</td>
<td>3.67</td>
<td>0.64</td>
<td>0.23</td>
<td>0.40</td>
<td>0.87</td>
<td>0.21</td>
<td>0.90</td>
<td>2.83</td>
<td>0.50</td>
</tr>
</tbody>
</table>
Results and Discussion

Sequence Attributes

The aligned matrix of 1263 bp of mtDNA sequence for 306 individuals (five outgroup, 301 ingroup) provided a total of 420 variable characters, of which 293 were potentially parsimony informative. Among the 301 *Ceuthophilus* ingroup taxa sequenced for this project, many share identical mtDNA sequences and hence the dataset was reduced to 182 unique haplotypes for phylogenetic analysis. These 182 unique haplotypes were divided among two subgenera, *Geotettix* and *Ceuthophilus*. Fifty-nine unique haplotypes were found in members of the subgenus *Geotettix* and 123 unique haplotypes were found among members of the subgenus *Ceuthophilus*.

Among ingroup taxa in the subgenus *Geotettix*, uncorrected sequence divergence ranged from 0.0 to 2.3% (mean = 1.1%). Whereas for the subgenus *Ceuthophilus*, uncorrected sequence divergence are higher and ranged from 0.0 to 9.3% (mean = 2.7%). These ranges and means are concordant with the pattern seen in the phylograms (Figures 9, 10) of these subgenera.

Phylogenetic Analyses and Patterns of Genetic and Phylogenetic Diversity

Neighbor-joining analyses of each subgenus produced well-supported phylogenies (Figures 11, 12). The monophyly of both the *Geotettix* and *Ceuthophilus* subgeneric clades were well supported by bootstrapping (88% for the *Ceuithophilus* subgeneric clade, 100% for the *Geotettix* subgeneric clade). The Bayesian mixed-model analysis of the reduced taxon dataset (47 terminal taxa) also produced a strongly supported phylogenetic hypothesis (Figure 13) with all except six nodes supported by ≥90% posterior probability. As expected, the subgenera *Geotettix* and *Ceuthophilus* are reciprocally monophyletic in this analysis. Several salient findings emerged from our analysis, which are discussed below.

First, the neighbor-joining haplotype phylogram for the subgenus *Ceuthophilus* shows deeper internal branches leading to clades than does the haplotype phylogram for the subgenus *Geotettix* (Figures 9, 10). Branch lengths in the Bayesian tree produced from the reduced dataset (Figure 13) also show deeper (older) genetic structure in the subgenus *Ceuthophilus* than that found in the codistributed subgenus *Geotettix*.

At least some crickets in the subgenus *Ceuthophilus* are known to be more vagile than those in the subgenus *Geotettix* (Taylor et al. 2003, 2004, 2005). Taylor et al. (2003, 2004, 2005) found that *Ceuthophilus* (*C.* secretus) individuals traveled more than 100 m from their cave entrance to forage at night, whereas *C. (G.) cunicularis* were never observed leaving their caves. Similarly, Veni and Associates (2006) observed only one instance of *C. (G.) cunicularis* exiting a cave during their extensive monitoring at Camp Bullis (Bexar County, Texas). Hence, we predicted that patterns of genetic structure for these two subgenera would differ with the cave-limited subgenus *Geotettix* showing higher levels of genetic structure than those in the subgenus *Ceuthophilus*. Instead, the genetic data differ from our prediction, with the subgenus *Ceuthophilus* having higher levels of genetic variation (uncorrected p-distance 0.0 - 9.3%) than found in *Geotettix* (uncorrected p-distance 0.0-2.3%). This pattern is also borne out in the phylogenetic analyses, which show that across the same geographic area, the subgenus *Ceuthophilus* has longer internal branches, and hence longer times since diversification than the subgenus *Geotettix*. One possible interpretation of this pattern is that subgenus...
Figure 9. Ceuthophilus (Ceuthophilus) spp. neighbor-joining tree, constructed from uncorrected p-distances, showing the genealogical relationships of haplotypes. Branch lengths are proportional to uncorrected p-distance as indicated by the scale bar. Samples are given as: Site Name, County-Specimen Numbers. Bold letters on left correspond to letters used in text.
Figure 10. *Ceuthophilus (Geotettix)* spp. neighbor-joining tree, constructed from uncorrected p-distances, showing the genealogical relationships of haplotypes. Branch lengths are proportional to uncorrected p-distance as indicated by the scale bar. Samples are given as: *Site Name, County-Specimen Numbers*. Bold letters on left correspond to letters used in text.
Figure 11. *Ceuthophilus* (*Ceuthophilus*) spp. haplotype neighbor-joining bootstrap consensus tree, constructed from uncorrected p-distances. Numbers above branches are support from 1000 bootstrap replicates (only values >50% are shown). Samples are given as: Site Name, County-Specimen Numbers. Bold letters on right correspond to letters used in text. Inset shows portion of tree displayed on this page (in white), remaining tree in insect (in gray) shown on following page as a continuation of this figure. (This figure continued on following page.)
Figure 11 (continued). *Ceuthophilus* (*Ceuthophilus*) spp. haplotype neighbor-joining bootstrap consensus tree, constructed from uncorrected p-distances. Numbers above branches are support from 1000 bootstrap replicates (only values >50% are shown). Samples are given as: *Site Name, County-Specimen Numbers*. Bold letters on right correspond to letters used in text. Inset shows portion of tree displayed on this page (in white), remaining tree in insect (in gray) shown on preceding page as the first part of this figure.
Figure 12. *Ceuthophilus* (*Geotettix*) spp. haplotype neighbor-joining bootstrap consensus tree, constructed from uncorrected p-distances. Numbers above branches are support from 1000 bootstrap replicates (only values >50% are shown). Samples are given as: Site Name, County-Specimen Numbers. Bold letters on right correspond to letters used in text.
Figure 13. Phylogenetic, cluster, and principal component analyses (PCA) of Ceuthophilus species color-coded by subgenera. A: mixed-model Bayesian analysis of combined COI and ND5 sequences (1263 bp), all clades with posterior probabilities >0.9, except those marked with a circle; B: single lineage cluster using Euclidean distances among principal component scores reduced from 17 measurements of females, numbers above branches refer to bootstrap percentages; C: strict consensus of nine most parsimonious trees (L=72.810) resulting from the analysis of 19 discrete and 16 continuous characters from males and females; D: 2-dimensional plots of PCA factors reduced from female measurement data of all Ceuthophilus specimens studied (>85% of explained variation) grouped by subgenera.
*Ceuthophilus* taxa have been isolated in particular caves or sets of caves for a longer period of time than the codistributed subgenus *Geotettix* taxa.

Second, in the neighbor-joining haplotype tree for the subgenus *Ceuthophilus*, different individual crickets from the same geographic areas (cave localities) fall out in multiple divergent haplotype groups (Figures 9, 11). For example, individual crickets from three distinct (divergent) haplotype groups are found in Behrens Grotto, Mason County (A, C, E), and Writing on the Rocks Cave, Edwards County (A, E, H). Haplotypes from two genetically distinct (divergent) groups are found in Porcupine Pit, Mason County (A), Tall Tales Cave, Bexar County (D, G), and Lamm Cave, Travis County, Texas (*C. [C.] secretus*, B). For the subgenus *Geotettix*, fewer caves harbor multiple divergent haplotype lineages (Figures 10, 12). In the case of MARS Shaft, Bexar County (*C. (G.) cunicularis*, Q) and Writing on the Rocks Cave, Edwards County (K, L), the co-occurring haplotype clades are not very divergent and likely do not represent unique species. Even so, Swift Cave, Mason County contains two described species in the subgenus *Geotettix* (*C. polingi, C. cunicularis*), and Tall Tales Cave, Bexar County contains two distinctly divergent haplotype clades (*M, C. cunicularis*). Thus, with notably fewer exceptions than in the *Ceuthophilus* subgenus, individual *Geotettix* from the same cave typically fall out into the same haplotype clade within the subgeneric haplotype tree. When both subgenera are considered together, however, there are a surprising number of caves at which more than two haplotype clades co-occur (Figure 15). Some of the caves with only one clade represented are undersampled, with additional clades present, but not represented in our samples.

Previous work has suggested that most caves held two *Ceuthophilus* cricket species, one from the subgenus *Ceuthophilus* and the other from the subgenus *Geotettix*. However, within both subgenera, undescribed species likely exist. Our phylogenetic data confirms this hypothesis and these data are consistent with the species level taxonomy of these cricket subgenera being higher than current taxonomy indicates. Both subgeneric phylogenetic trees show multiple divergent lineages that have likely been on their own evolutionary trajectories for a considerable period of time. Furthermore, the phylogenetic analysis of haplotypes identifies several instances in which individual crickets from the subgenus *Ceuthophilus* collected from the same locality fall out in two or three different haplotype clades, which suggests that individual caves hold more than the traditionally hypothesized two species. For example, Behrens Grotto, Mason County, has one genetic lineage of *Geotettix* (*C. (G.) polingi*) and three lineages of the subgenus *Ceuthophilus* (A, C, E). Whether these genetic lineages are actually species will require further study, but certainly this pattern
of genetic diversity also suggests, in general, that the genus *Ceuthophilus* may hold more diversity than that described by Hubbell (1936).

Lastly, our visual analysis of the phylogenetic tree mapped on geography, performed using Mesquite’s Cartographer module (Maddison and Maddison 2006a, 2006b), suggests that geographically proximate individuals share more similar mtDNA sequences than do geographically distant specimens (Figure 16). Hence geographic distance may be related to patterns of genetic structure in these two cricket subgenera. For example, two biogeographic patterns stand out in the genetic analysis: 1) a major discontinuity just north of the Colorado River, a probable barrier to distribution, and 2) a discontinuity in far west Texas where karst rocks are not exposed and the surface is extremely arid.

In the subgenus *Ceuthophilus*, the discontinuity to the north of the Colorado River is evident when examining the relationship of clades *C. (C.) secretus*, F and E (Figures 9, 17). The first two clades occur only north of the discontinuity, in northern Travis, Williamson and Coryell counties, and the sister group to F, clade E, occurs only south of the discontinuity. However, clade B (Figures 9, 17) is inconsistent because it has representatives that cluster together from both sides (in particular, Lamm Cave). This inconsistency could arise from a history of multiple founders crossing the barrier at different times, causing one population (or a putative species) from Lamm Cave to cluster with others north of the divide and another to cluster with those that otherwise occur south of the divide. Another explanation is that clade B is all that remains of an older lineage that was widespread prior to the development of the Colorado River as a formidable barrier to distribution. A third possibility is that the historic path of the Colorado River swung as far north as Lamm Cave such that at some times in history that area was south, or very close to the Colorado River.

For *Geotettix* subgenus, clade N is entirely north of the Colorado River, while all the other clades, with the exception of one population, occur south of the Colorado River (Figures 10, 18). Again that exception is Lamm Cave in Travis County. The same possibilities exist for this inconsistency, and both subgenera deserve a higher density of sampling in the area to help clarify their evolutionary history. Other cave taxa show a similar pattern of species break at the Colorado River. The aquatic asellid isopod *Lirceolus* spp. occurs only south of the Colorado River, and the aquatic cirroniid isopod *Cirolanides texensis* occurs only south of the Colorado River with one exception, Longhorn Caverns (Krejca 2005). The troglobitic harvestman genus *Chinquipellobunus* occurs only south of the Colorado River, and the troglobitic harvestman *Texella reddelli* occurs only south while closely related *Texella* congeners occur north of this divide. Undoubtedly a more thorough search of

---

Figure 16 (on following page). Phylogenetic and principal component analyses (PCA) of the subgenus *Ceuthophilus* (*Geotettix)*. Genetic lineages are grouped based on major clades, which were color-coded and designated by a different symbol. A: cropped Bayesian analysis of COI+ND5 sequences; B: 2-dimensional plots of PCA factors reduced from female measurement data of specimens studied by genetic groups following part A of this figure; C: plot of phylogenetic tree over map of south-central USA using the information of geographical coordinates of terminal taxa, grouped following part A of this figure; D: 2-dimensional plots of PCA factors reduced from female measurement data of specimens studied grouped by pseudosternite shape; E: 2-dimensional plots of PCA factors reduced from female measurement data of specimens studied grouped by aspect of setae row on plantar surface of metatarsomere I of females and males. Geological/aquifer map in C adapted from Miller (1998).
Figure 17. Distribution of clades sampled for *Ceuthophilus* (*Ceuthophilus*) spp. Clades recorded from a single site are indicated by small circles (*C. [C.] longipes*, *C. [C.] conicaudus*, F, J); clades recorded from two sites are indicated by lines (*C. [C.] secretus*, I); clades recorded from three or more sites are indicated by minimum convex polygons (A, B, C, D, E, G, H).
Figure 18. Distribution of clades sampled for *Ceuthophilus* (Geotettix) spp. Clades recorded from a single site are indicated by small circles (*C. [G.] carlsbadensis*, K, S, T, Q, Q2); clades recorded from two sites are indicated by lines (M, O, P); clades recorded from three or more sites are indicated by minimum convex polygons (*C. [G.] cunicularis*, *C. [G.] polingi*, L, N).
the distribution of cave taxa would reveal more examples of the pattern. A phylogenetic analysis of approximately 18 species of troglobitic *Cicurina* spiders across central Texas did not yield any examples of populations that occurred on both sides of the Colorado River, though this hypothesis was not explicitly tested or discussed (Paquin and Hedin 2004). In *Eurycea* salamanders, the Colorado River represents the most basal node in the phylogenetic tree, dividing the northern group from all other lineages (Chippindale et al. 2000). In *Plathodon* salamanders, phylogenetic analyses showed a major phylogenetic break several kilometers north of the Colorado River, and hypothesized that these populations immediately north of the river clustered with southern populations because they occurred on terrace deposits that were located south of the river until the Pleistocene (Baird et al. 2006). Our data do not perfectly fit this model because our populations north of the river are not on terrace deposits.

Regarding the discontinuity of populations in west Texas and Mexico, both subgenera show similar but not totally consistent patterns. For the subgenus *Ceuthophilus*, a deep branch separates one population from 400 Foot Cave (Clade J) from the rest of Texas and some of New Mexico. This partially correlates with the Pecos River as a barrier, except that Amazing Maze Cave population clusters with others from east of the Pecos (Figure 17). Possibly in the farther upstream reaches of the Pecos, near Amazing Maze Cave, the River more frequently runs dry and does not pose a significant barrier to migration. As in the example at Lamm Cave, above, 400 Foot Cave also has a population that clusters with Clade C, probably a result of multiple invasions, historic widespread species, and changes in past climactic conditions that may have taken down present-day barriers.

In the subgenus *Geotettix*, there are deep branches separating isolated species in New Mexico and Mexico [(*C. (G.) carlsbadensis* and Clade T] corresponding to the desert in far west Texas and the Rio Grande (Figure 18). As in the previous discussion, however, there are some inconsistencies with the Rio Grande as a barrier because one of the two Mexico populations examined, *C. polingi* in Cueva de La Azufrosa, clusters with those from Uvalde and Mason counties. Molecular phylogenetic analysis of aquifer isopods in west Texas and northern Mexico also showed deep splits at the Pecos and Rio Grande rivers (Kreja 2005), and distribution breaks of aquifer asellid isopod species (Lewis 2001) occur at the non-cavernous region between the Carlsbad Cavern, New Mexico area and west Texas (white space in west Texas, Figure 17).

Our analysis shows that cave cricket genetic distance is dictated by geographic distance, but discussion herein demonstrates that major barriers present in other cave taxa are also shared by crickets. The common occurrence sympatric lineages at the same caves complicates our analysis, and in some areas, such as those immediately adjacent to large barriers (Colorado, Pecos, Rio Grande rivers), more sampling will be needed to clarify these relationships. Additionally, further post-hoc hypothesis testing specific to the presumed barriers, such as with a parametric bootstrap analysis (similar to those done in Kreja 2005) and Mantel tests of genetic distances versus great circle geographic distances, could improve our understanding of the role those barriers play. Such analyses will be incorporated into our study prior to final publication in a peer-reviewed journal.

**Morphological species**

Morphological species concepts were mostly based on the shape of the sclerotized pseudosternite, which Hubbell (1936) used extensively in his taxonomic revision of the genus *Ceuthophilus* and considered that “The modifications of this structure are of the utmost taxonomic value in the study of the genus.” Based on the study of the pseudosternite morphology, the identity of *C. (C.) secretus* and *C. (G.) carlsbadensis* was confirmed. One male specimen previously identified as
C. (C.) conicaudus (#258) keys to this species using Hubbell (1936), but the arch dorsum of the pseudosternite in the specimen is a bit broader and less projected than in the specimen figured by Hubbell (1936) and has two ventral subdistal spurs of metatibiae. More than 95% of C. (C.) conicaudus specimens (n>80) studied by Hubbell had a single subdistal spur, which he considered a relatively constant character and used it as a main differentiating character of C. (C.) conicaudus and C. (C.) secretus, 98% of the latter having two subdistal spurs. However, this male specimen and sympatric female (#259) lack the main reliable diagnostic characters of C. (C.) secretus: (1) pseudosternite arch internally with two sclerotized tumid lobes (Figure 19F); (2) paraprocts with short and stout spines (Figure 19C); and female ovipositor with six apical teeth. The specimens were collected in Carlsbad Cavern, Eddy County, New Mexico, and specimens of C. (C.) conicaudus from caves in Dark Canyon, just north of Carlsbad Caverns National Park and 20.1 km southwest of the city of Carlsbad were studied by Hubbell (1936) when he described the species as new. In addition, several other researchers studied crickets in southeastern New Mexico, and these, too, refer to C. (C.) conicaudus (Barr and Reddell [1967], Campbell [1976], Cokendolphr [2001], Cokendolpher and Polyak [1996, 2004], Northup and Kuper [1987], and Northup et al. [1993]; see Lavoie et al. [2007] for a review of literature). Therefore, this lineage is herein referred to as C. (C.) conicaudus, despite the incongruencies.

Other morphological species belonging to the subgenus Ceuthophilus (Geotettix) were herein identified as C. (G.) cunicularis, and C. (G.) polingi, with clade S also seeming markedly unique (see discussion below). Male specimen #220 keys unequivocally to C. (G.) cunicularis, a species described from Texas (Bexar, Hays, and Travis counties). Hubbell (1936) relates this species to C. (G.) polingi and Ceuthophilus (Geotettix) umbratilis, but distinguishes it because of the male terminalia. Although all three species mentioned share a similar median longitudinal crest (Figure 19A-F), only C. (G.) cunicularis has a median triangular projection on the margin of dorsal rim, recurvate in lateral view (Figure 19G, I-L). Interestingly, this basic pseudosternite shape is also found in almost all genetic lineages of Ceuthophilus (Geotettix) (Figure 16D). However, males of all these other lineages do not key to C. (G.) cunicularis basically because the plantar surface of their metatarsomere I does not have a complete row of setae along most of its length (Figure 7I, 16E). The only exception is lineage M, which does have a complete row of setae (Figure 7J), but the projection on the margin of dorsal rim is much broader and less triangular than that described for C. (G.) cunicularis. Furthermore, although all these lineages do share the same basic plan, there is quite a bit of intergrading variation of some key characteristics in the male pseudosternite; such as, in the shape and size of the dorsal rim process, the degree to which this process is reflected upward, and the height and shape of the dorsal crest. This variation is further made difficult to quantify due to subjective artifacts as the positioning of the sclerite. Additionally, the male specimen from Comal County has the profemur anteroventral carina (Figure 7G: PAV) with 2-3 spurs and mesofemur anteroventral carina with five spurs, consistent with the original description of C. (G.) cunicularis. Therefore, only these specimens from Comal County are henceforth referred to as C. (G.) cunicularis, while other members of the Q lineage (including Q2) are not.
A number of the divergent clades in the mitochondrial haplotypes trees may represent undescribed and/or cryptic species. For example, molecular lineage S may be morphologically and geographically distinct enough for it to be considered a new species within the subgenus Geotettix. Lineage S is separated geographically and geologically from other Ceuthophilus (Geotettix). We recorded this species only from 400 Foot Cave, outside the Edwards Plateau in the Glass Mountains of Brewster County, Texas. Other specimens collected in this cave are also very interesting and fall in lineages I and J of Ceuthophilus (Ceuthophilus), but unfortunately no adult specimens were collected of lineage I and only 2 adult females were collected from lineage J. Although, not highly genetically (Figure 13A) or morphometrically (Figure 13B) divergent from other Ceuthophilus (Geotettix) studied herein, the male genital characters of lineage S are quite distinct. Lineage S has very distinct subgenital plates with apicolateral projections very widely separated (Figure 19O), a pseudosternite arch with a dorsum lacking the median longitudinal crest, and a dorsal rim with a very broad subquadrat median process which is not distinctly upturned like other cunicularis-like pseudosternites (Figure 19H). Finally, it is worth noting that Cokendolpher and Polyak (1996) indicate that C. (G.) carlsbadensis is known from Brewster, Culberson, and Jeff Davis counties in Texas – it is possible that some of the collections to which they refer are attributable to our lineage S. More material from this area should be subject to morphological analysis to assess the range of structural variation before the taxonomic status of lineage S is finally determined.

Finally, some specimens belonging to the genetic lineage R were identified as C. polingi based on the distinctive pseudosternite morphology (Figure 19G, N). Ceuthophilus (G.) polingi was described based on two males from Davis County, Texas. This species is therefore herein recorded for the first time from Mason and Uvalde counties, Texas and Coahuila, Mexico. Strangely, a very large male specimen (#454) grouping with the remaining C. (G.) polingi (Figure 16A) from approximately the middle of this range (Uvalde County, Texas, see Figure 16C), not only has a cunicularis-like pseudosternite, but also lacks the complete row of setae along the plantar surface of the metatarsomere I (Figure 7I) diagnostic of C. (G.) polingi.

### Congruence of DNA-based phylogeny and morphological data

The Bayesian phylogenetic analysis is based on DNA-sequences (Figure 13A), parsimony analysis of combined continuous and discrete morphological characters (Figure 13B), and cluster analysis based on distances of the principal component scores (Figure 13C). Overall, there is almost no congruence in the relationships suggested by each methodology and dataset. The only striking similarity is that both the molecular and combined morphology datasets generally support the two subgenera as distinct. They are monophyletic sister groups in the molecular analysis (Figure 13A), and one morphology tree shows Geotettix as a monophyletic lineage of Ceuthophilus (Figure 13B), while the other configures Geotettix as paraphyletic and Ceuthophilus as polyphyletic (Figure 13C). Although, the PCA score cluster resulted in a non-congruent hypothesis with the other datasets (Figure 13C), the 2-dimensional plots were successful in separating members of the two different subgenera (Figure 13D) and, considering the variability of some key characters (for example, the variation of characteristics in the male pseudosternite, discussed earlier), may be generally more useful when comparing morphology to genetics.

In the Ceuthophilus (Ceuthophilus) lineage, the male genitalia of relatively fewer specimens were studied because of the lesser availability of adult forms. Although the clades recovered by the Bayesian analysis had good statistical support (based on posterior probabilities), the three defined multi-lineage groups (Figure 20A, red circle, blue square, and green diamond) were not supported by the parsimony analysis of morphological characters (Figure 13B) and were not distinctly grouped in
the 2-dimensional PCA plots (Figure 20B). Only one of the multi-lineage groups (C. (C.) conicaudus + G + H) (Figure 20A) was somewhat similar to the strict consensus of nine most parsimonious trees (Figure 13C). However, the PCA analysis shows a large separation of lineage J from other members of the subgenus, as does the DNA analysis (Figure 20). Furthermore, the morphometric data based on females supports the grouping of C. (C.) secretus + lineages D + E (Figure 20B), which was not recovered with the DNA analysis (Figure 20A). Both males and females of these three lineages have the distinguishing characters abovementioned for C. (C.) secretus. The DNA analysis also supports the grouping of lineage F within the C. (C.) secretus lineage. However, males of lineage F were not available for study, and although geographically consistent with C. (C.) secretus distribution, the female studied bore five teeth in the ventral valve instead of the characteristic six teeth. Aberrant females of C. (C.) secretus with only five teeth were previously studied by Hubbell (1936). The lack of information on the male genitalia and the unusual female genitalia of lineage F probably caused it to not group with C. (C.) secretus + D + E in the morphological analysis (Figure 13B).

Members of Ceuthophilus (Geotettix) are differentiated morphologically from other Ceuthophilus subgenera by its usually smaller size and more compact shape (shorter legs), ventral carina of the metafemur with several more denticulations, and a distinctive putative synapomorphy of males having the abdominal tergite VIII strongly produced, usually concealing the little-produced tergite IX, with its apex usually tumid and round (Figure 7A) (Hubbell 1936). Another synapomorphy recovered by this study is the presence of a denticle on the anterior genicular lobe of the metafemur in all studied Ceuthophilus (Geotettix) members. All studied specimens fall into what Hubbell (1936) defined as the silvestris group of species. Except for C. (G.) carlsbadensis, the phylogenetic history
based on DNA sequences shows very small divergences between members of this closely related
group, which contributed to low statistical branch support of some multi-lineage groups (Figure 13A). As abovementioned, most studied males of this group have a *cunicularis*-like pseudosternite, with the exception of lineage S and *C. (G.) polingi* which appear to be confidently delimited morphological species. However, both of these lineages do not show strikingly longer branch lengths than other lineages within this clade. Whether both species are indeed genetically separated from the other *cunicularis*-like lineages is impossible to say with such a small taxon sampling in this analysis and lacking statistical support for the relevant multi-lineage clades in the phylogeny.

Finally, it is interesting to note a couple of minor incongruencies between the resulting DNA-sequence topology (Figure 16A) and morphometric analysis (Figure 16B) of *Ceuthophilus (Geotettix)*. A male specimen from Bexar County included in lineage M (#240) and sympatric female (#219), based on DNA sequences, appear to be more related to lineages K, L, and O (blue triangle group). However, the female is much more similar morphometrically to the group including *C. (G.) cunicularis*, P and Q (upside-down green triangle), both males and females have a complete row of plantar setae on metatarsomere I like *C. (G.) cunicularis* (Figure 16E), and their distribution seems more congruent with the green group (Figure 16C). Conversely, the male specimen (#116) belonging to Q2 and positioned in the upside-down green triangle seems more similar to members of the group containing lineages K, L, and O (blue triangle group). Although this specimen from Kerr County is not distinctly dissimilar in genitalic characters from males of the Q lineage, the sympatric female (#114) is morphometrically very distinct from most other *Ceuthophilus (Geotettix)* studied (Figure 16B). The metafemur armature is very well-developed similar to the female specimen from lineage O. Similar to the blue group, males lack the complete row of and females have a few basal setae on the plantar surface of metatarsomere I (Figure 16E).

Concerning the characters of presence and extent of the ventral row of setae on the plantar surface of metatarsomere I (Figure 7I, J), Hubbell (1936) considered the presence of these setae (*Type B* armature) a “retention by the adult of a condition universal among early instars, but which in most species is lost in some pre-adult stadium.” He further diagnosed the *silvestris*-group based on this *Type B* tarsal armature, with the exception of *C. (G.) carlsbadensis*, which normally has tarsus *Type A* (Hubbell 1936). Although he did not mention this character or specify the sexual dimorphism in the species descriptions of this group, it is used in both male and female taxonomic keys to species of *Ceuthophilus*. It is reported here as many additional lineages of the *silvestris*-group, which share a similar pseudosternite shape with *C. (G.) cunicularis*, but like *C. (G.) carlsbadensis*, their males lack setae on the plantar surface, and in all cases (except in lineage L) the females do have basal setae (Figure 16E). Clearly, the morphological character set used by Hubble (1936) (and in the present study) is inadequate to explain species-level differences among lineages. A search for additional characters, including the use of characters the male phallus (not examined in this study) should be undertaken, and future taxonomic decisions within the genus *Ceuthophilus* also should take genetic data into consideration.

**Conclusions**

Both *Ceuthophilus* subgenera show considerable geographic and phylogenetic structure, as well as significant morphological variation. The longer branch lengths and morphological variation found in the subgenus *Ceuthophilus* suggests less genetic communication among populations in this subgenus relative to the subgenus *Geotettix*, a finding that is surprising and contrary to our expectation, because members of the subgenus *Ceuthophilus* commonly forage outside of caves in central Texas. In
addition, some caves have multiple mitochondrial lineages in them that possibly represent cryptic species, implicating that diversity is higher than previously expected. We identified 22 different clades (C. [Ceuthophilus] spp. [10 clades]: A, B, C, D, E, F, G, H, I, J, C. [C.] secretus; and C. [Geotettix] spp. [12 clades]: K, L, M, N, O, P, Q, Q2, R, S, C. [G.] cunicularis, C. [G.] polingi) in Texas caves which belong to the genus Ceuthophilus and are on independent evolutionary trajectories. Some of these clades may represent undescribed or cryptic species, and thus suggests that biotic diversity in Ceuthophilus is higher than current species level taxonomy indicates. These data point towards a need for additional conservation attention in a variety of Texas cave systems, and additional research, including additional genetic and traditional taxonomic work, are needed to make educated conservation decisions regarding the diverse fauna found in these cave systems.

Acknowledgements

Many people helped make this report possible. In addition to the innumerable landowners and land managers, both private and government (including Texas Parks and Wildlife, The Nature Conservancy, The Texas Cave Management Association, U.S. Fish & Wildlife, Fort Hood, Camp Bullis, Fort Leonard Wood, Caverns of Sonora, etc.) and the cavers and other who created maps of caves that facilitated our visits, or helped out in phone calls or conversations in finding caves and helping us gain access, the following individuals made special efforts to help us acquire the samples needed for this study: Peter Sprouse, Kristopher Megahan, N. Parker, Rob Meyers, Peter Sprouse, A. Gluesenkamp, Vivian Loftin, Cindy A. Lee, Justin Shaw, Alan King, Jim Kennedy, M. Sanders, K. O'Connor, J. Chenowith, Alan King, Wayne Peplinski, Don Evan Selby, Allan Cobb, C. Phillips, C. Pekins, Bill Sawyer, Michael Cunningham, Michael E. Slay, Kurt L. Helf. Many of the above individuals participated in multiple trips and spent long hours driving between sites, crawling through caves, and fending off rattlesnakes. We thank Don Selby, who spent time trying to find Ceuthophilus secretus in the crawlspaces of homes in Dallas (the type locality). JoAnn Jacoby reviewed an earlier draft of this report. We apologize to those we’ve undoubtedly overlooked

Literature Cited


40


Schneider, S., D. Roessli, and L. Excoffier. 2000. Arlequin ver. 2.000: A software for population genetics data analysis. Genetics and Biometry Laboratory, University of Geneva, Switzerland.


Appendix 1. Morphological discrete characters and states.

Male terminalia:
1. Abdominal tergite VIII: (0) well produced farther than tergite IX (Figure 4A); (1) not produced farther than state IX
2. Subgenital plates: (0) fused basally and apicolateral portions produced (Figure 4B); (1) divided basally by sulcus and apicolateral portions not distinctly produced
3. Epiproct shape: (0) triangular (Figure 4C); (1) subquadrate
4. Paraprocts sculpturing: (0) without stout spines; (1) with short and stout spines (Figure 4C)
5. Pseudosternite dorsum: (0) without longitudinal crests (Figure 4D); (1) with median crest; (2) with pair of parallel crests (Figure 4E)
6. Pseudosternite dorsal rim of arch: (0) without processes (Figure 4D); (1) with median process
7. Pseudosternite median process of dorsal rim of arch: (0) triangular; (1) broad, quadrangular
8. Internal wall of arch: (0) without folds; (1) with pair of tumid folds (Figure 4F)

Female thorax and appendages:
9. Pronotal recumbent setae: (0) absent; (1) present
10. Metafemoral anterior genicular lobe denticle: (0) absent; (1) present (Figure 4H)
11. Metafemoral posterior genicular lobe denticle: (0) absent; (1) present
12. Metatibial subdistal ventral spurs: (0) 2:1, (1) 2:1:1; (2) 1:1
13. Metatarsomere I chaetotaxy: (0) absent or few basally (Figure 4I); (1) single row throughout most tarsomere (Figure 4J)

Female ovipositor:
14. Ovipositor ventral valve number of teeth: (0) 5 (Figure 5K); (1) 6

Female spur and denticulation counts:
15. Profemoral AV carina spurs (Figure 4G: PAV): (0) absent; (1) 1-7; (2) 9; (3) 11; (4) 23
16. Mesofemoral ventral carinae spurs: (0) absent; (1) 1-12; (2) 17-24; (3) 56
17. Metafemoral ventral carinae denticulations (Figure 4H: VCD): (0) absent; (1) 1-120; (2) 141-200; (3) 242; (4) 497
18. Metafemoral dorsal denticulations (Figure 4G: DD): (0) absent; (1) 1-25; (2) 38; (3) 60-93; (4) 103-110; (5) 129
19. Metatibial basal AD carina denticulations (before basal spur): (0) 7; (1) 9-10; (2) 12-14; (3) 16-17; (4) 19-21
Appendix 2. List of morphometric characters used to calculate the continuous ratios.

1. Eye area (Figure 5B)
2. Interocular distance (Figure 5B)
3. Fastigium distance to mesal margin of antennal fossa (Figure 5C)
4. Maxillary palp distal segment length (Figure 5A)
5. Profemur length (Figure 5D)
6. Profemur AV1 spur length (Figure 5E)
7. Mesofemur posterior genicular spur length (Figure 5F)
8. Metafemur length (Figure 5G)
9. Metafemur width (Figure 5G)
10. Metatibia dorsal subdistal spur length (Figure 5H)
11. Metatibia AD calcar length (Figure 5H)
12. Metatarsomere I length (Figure 5H)
13. Metatarsal claw length (Figure 5I)
14. Cercus length (Figure 5J)
15. Ovipositor ventral valve length (Figure 5K)
16. Ovipositor ventral valve armed region length (Figure 5K)